



2-butene (5a). Although some coupling was observed in diethyl ether the reaction was too slow to be useful. The corresponding bromide (5b) on the other hand gave diene 6 in close to quantitative yield based on the vinylic lithium 2, which was formed in 70–90 % yield from the bromide 1 (*vide infra*).

The homo-coupling product, 2,5-dimethyl-2,4-hexadiene, amounted to less than 2 % of the final reaction mixture and no trace of the product anticipated from an allylic rearrangement (*cf.* Ref. 9) of the electrophile 5b was present as judged by capillary GC.

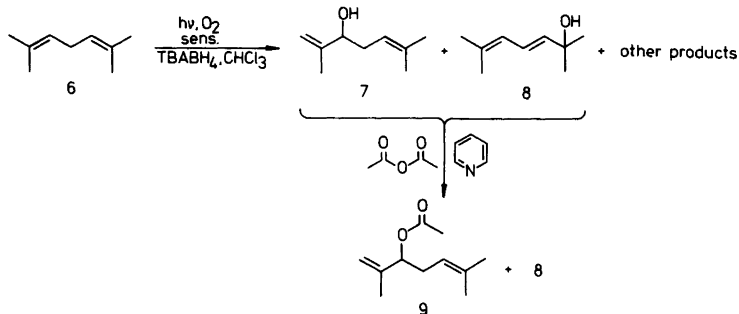
Coupling of the vinylic lithium reagent 2 with the allylic bromide 5b was also tried in the absence of Cu(I) (*cf.* Ref. 10). In this case a very slow formation of the cross coupling product was observed. In the presence of 0.5 equivalents of TBACu(I)Br<sub>2</sub> (3) on the other hand, the reaction rapidly went to completion in a close to quantitative yield calculated on the amount of vinylic lithium compound 2 formed. This amount was estimated by quenching with water and subsequent GC determination of the ratio between vinylic bromide 1 and 2-methylpropene. This means that both the vinylic groups of cuprate 4 can be transferred under mild conditions (*cf.* Ref. 11).

Our route to the mealybug pheromone (9) is outlined in Scheme 2. Photooxidation of 2,6-dimethyl-2,5-heptadiene (6) in chloroform in the presence of TBABH<sub>4</sub><sup>2</sup> and subsequent treatment of the crude reaction mixture with acetic anhydride and pyridine selectively acetylated the secondary alcohol 7. The desired acetate 9 was then easily separated by liquid chromatography. The alcohols 7 and 8 could also be separated by chromatography prior to acetylation but such a

procedure offers no advantage. Due to unfavourable product distribution and secondary product formation the overall yield of the racemic pheromone 9 was only ≈10 %. This result is in accord with a report on a conventional photooxidation of diene 6 which appeared after the initiation of this work.<sup>12</sup> In spite of the low yield, the described method makes the racemic pheromone of the Comstock mealybug available by simple operations.

## EXPERIMENTAL

<sup>1</sup>H NMR spectra were recorded in CDCl<sub>3</sub> on a Bruker WP 200 or a JEOL PMX-60 spectrometer. A Finnigan model 4021 connected to an INCOS data system was used to record GC-MS spectra. Analytical GC was performed on a PYE 204 instrument with an FID detector using fused silica capillary columns. Liquid chromatography was performed with Merck silica gel 0.040–0.063 mm dry packed in 2.54 cm i.d. columns unless specified. The solvent, light petroleum b.p. 40–60 °C with increasing amounts of ethyl acetate (0, 0.63, 1.25, 1.88, 2.5, 3.75, 5.00, 7.5, 10, 15, 20, 30, 40, 60, 80 %) was delivered with a metering pump at a rate of 60 ml/min. A "poor man's" gradient mixer was arranged as follows: The suction tube of the pump was placed in a stirred vessel initially containing pure light petroleum and portions of solvent of increased polarity were successively delivered to the surface of the liquid in the vessel by a stoppered dropping funnel. All fractions (30 ml) were examined by TLC. TLC was performed on silica gel (Merck 60, precoated aluminium foil) eluted with 20 % ethyl acetate in light petroleum. Spraying with vanillin in ethanol containing sulfuric acid and heating to 140 °C visualized the compounds. Irradiations were performed in



Scheme 2.

Pyrex vessels using a Rayonet reactor with 16 RPR 350 nm lamps, employing the procedure for photooxidation in the presence of TBABH<sub>4</sub> described earlier.<sup>2</sup> The coupling reactions were performed strictly under argon atmosphere and all transfers were made *via* a double-ended needle or a syringe.

**2,6-Dimethyl-2,5-heptadiene (6).**<sup>13</sup> 1-Bromo-2-methyl-1-propene (*I*)<sup>14</sup> (3.2 g, 23.7 mmol) in dry ethyl ether (30 ml) was slowly added to finely cut lithium metal (0.36 g, 2.1 eq., sodium content 4.25 %) in ethyl ether (60 ml) at -25 °C. The temperature was kept below -15 °C throughout the lithiation which was monitored by withdrawing samples that were quenched with water. The ratio of the starting bromide (*I*) and the formed 2-methyl propene was determined by GC with a 25 m SE-30 capillary column. The lithiation was complete after 2 h and the solution was transferred to a suspension of TBACu(I)Br<sub>2</sub> (*3*)<sup>8</sup> (5.26 g, 12.1 mmol) in ethyl ether (100 ml) at -70 °C. Then 1-bromo-3-methyl-2-butene (*5b*) (2.9 g, 19.4 mmol) in ethyl ether (20 ml) was added. The coupling reaction went to completion in a few minutes and was worked up by addition of a saturated aqueous solution of ammonium chloride at -70 °, extraction with ethyl ether, washing the ether with brine, drying (MgSO<sub>4</sub>) and filtering through silica gel. Careful removal of the solvent gave 2.4 g of a product which was shown by GLC to be 98.5 % pure (99 % based on the allylic bromide *5b*). Spectral data were consistent with those published earlier.<sup>13</sup>

**Photooxidation of 2,6-dimethyl-2,5-heptadiene (6).** Isolation of 2,6-dimethyl-1,5-heptadien-3-ol (*7*) and 2,6-dimethyl-3,5-heptadien-2-ol (*8*). 2,6-Dimethyl-2,5-heptadiene (*6*) (0.950 g, 7.66 mmol), TBABH<sub>4</sub> (2.00 g) and dodecane (0.264 g) as internal standard were dissolved in chloroform (90 ml) containing TBA-solubilized rose bengal (1.5 g rose bengal and 0.96 g TBABr/l). The reaction mixture was irradiated with oxygen bubbling through the solution and the disappearance of starting material was monitored by GC (25 m SE-54 capillary column). At 65 % conversion of starting material (75 min) the irradiation was stopped and additional TBABH<sub>4</sub> (0.5 g) was added. After 30 min the chloroform was removed *in vacuo*. Addition of potassium iodide (2.5 g), water (3 ml) and ethyl ether (25 ml) caused TBA-iodide to precipitate. After 2 h of stirring the precipitate was removed by filtration and washed with ethyl ether. Washing the ether with brine, drying (MgSO<sub>4</sub>) and solvent removal gave a red viscous residue which was adsorbed on silica gel (5 g) and chromatographed on a 33 cm column topped with neutral alumina (0.5 cm) to retain the dye. Fractions 32-34 contained

2,6-dimethyl-1,5-heptadien-3-ol (*7*) (0.117 g, 10.9 %) and fractions 36-40, 2,6-dimethyl-3,5-heptadien-2-ol (*8*)<sup>15,16</sup> (0.255 g, 23.8 %). Recorded spectral data for compound *7*: MS,<sup>4b</sup> IR<sup>3,5b,6</sup> NMR.<sup>3,5b,6</sup> Compound *8* <sup>1</sup>H NMR 200 MHz: δ 6.44 (1H, dd, *J* 10.8 and 15.3 Hz, =CH-CH=CH-), 5.81 [1H, additional allyl coupling, *J* 10.8 Hz, (CH<sub>3</sub>)<sub>2</sub>=CH-CH=], 5.71 [1H, d, *J* 15.3 Hz, (CH<sub>3</sub>)<sub>2</sub>OH- -CH=CH-], 1.78 [6H, s, additional allyl coupling, (CH<sub>3</sub>)<sub>2</sub>=CH-], 1.34 [6H, s, (CH<sub>3</sub>)<sub>2</sub>OH-], 1.41 variable (1H, s, -OH).

**2,6-Dimethyl-1,5-heptadien-3-ol acetate (9)** via selective acetylation of the crude photolysate of 2,6-dimethyl-2,5-heptadiene (*6*). The diene *6* (2.5 g) and TBABH<sub>4</sub> (5.2 g) were dissolved in chloroform (90 ml) containing TBA-solubilized rose bengal and the solution was irradiated (150 min) as described above. Additional TBABH<sub>4</sub> (1.5 g) was added and after 30 min the reaction mixture was worked up, using potassium iodide (6.7 g), water (10 ml) and ethyl ether (100 ml). After solvent removal pyridine (5 ml) and acetic anhydride (5 ml) were added to the crude reaction mixture. After 3 h at ambient temperature brine (25 ml) was added and the reaction mixture extracted with light petroleum (3×25 ml). Drying and solvent removal gave a pyridine smelling residue which was adsorbed on silica gel (10 g) and chromatographed on a 33 cm column. This gave 0.337 g (9.2 %) of the racemic pheromone *9* with spectral data in agreement with those published earlier.<sup>4-6</sup>

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## REFERENCES

1. Henrick, C. A. *Tetrahedron* 33 (1977) 1845.
2. Baeckström, P., Okecha, S., De Silva, N., Wijekoon, D. and Norin, T. *Acta Chem. Scand. B.* 36 (1982) 31.
3. Shono, T., Yoshimura, T. and Oda, R. *J. Org. Chem.* 32 (1967) 1088.
4. a. Bierl-Leonhardt, B. A., Moreno, D. S., Schwarz, M., Forster, H. S., Plimmer, J. R. and DeVilbiss, E. D. *Life Sci.* 27 (1980) 399; b. Bierl-Leonhardt, B. A., Moreno, D. S.,

- Schwarz, M., Forster, H. S., Plimmer, J. R. and DeVilbiss, E. D. *J. Chem. Ecol.* 8 (1982) 689.
5. a. Negishi, T., Uchida, M., Tamaki, Y., Mori, K., Ishiwatari, T., Asano, S. and Nakagawa, K. *Appl. Entomol. Zool.* 15 (1980) 328; b. Uchida, M., Nakagawa, K., Negishi, T., Asano, S. and Mori, K. *Agric. Biol. Chem.* 45 (1981) 369.
  6. Mori, K. and Ueda, H. *Tetrahedron* 37 (1981) 2581.
  7. Theis, A. B. and Townsend, C. A. *Synth. Commun.* 11 (1981) 157.
  8. Nilsson, M. *Acta Chem. Scand. B* 36 (1982) 125.
  9. van Mourik, G. L. and Pabon, H. J. J. *Tetrahedron Lett.* (1978) 2705.
  10. Linstrumelle, G. *Tetrahedron Lett.* (1974) 3809.
  11. Alexakis, A., Cahiez, G. and Normant, J. F. *Synthesis* (1979) 826.
  12. Carless, H. A. J. and Batten, R. J. *Tetrahedron Lett.* (1982) 4735.
  13. Zilenovski, J. S. R. and Hall, S. S. *J. Org. Chem.* 44 (1979) 1159.
  14. Braude, E. A. and Evans, E. A. *J. Chem. Soc.* (1955) 3324.
  15. Braude, E. A. and Coles, J. A. *J. Chem. Soc.* (1952) 1425.
  16. Smith, I. H. and Casida, J. E. *Tetrahedron Lett.* (1981) 203.

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