

Studies on a Galactan from Norwegian Spruce Compression Wood (*Picea abies* Karst.)

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A galactan, containing about 13 % uronic acid residues, was isolated from Norwegian spruce compression wood. Partial hydrolysis gave D-galactose and a number of neutral oligosaccharides, which were shown by paper chromatography and paper electrophoresis to belong to the same homologous series. The first member of this series was crystalline and identified as 4-O- β -D-galactopyranosyl-D-galactose. The neutral part of the polysaccharide must therefore be composed essentially of linear chains of 1:4- β -linked D-galactose residues.

It has been known for some time that compression wood on hydrolysis yields a much higher proportion of galactose than normal wood¹. In the present investigation a polysaccharide containing galactose residues was isolated from compression wood from Norwegian spruce (*Picea abies* Karst.) and characterised. Acetone-extracted, milled compression wood, on extraction with hot water, yielded only a small fraction (A). However, a large amount of polysaccharide was isolated from the solution obtained on delignification of the wood meal with chlorite. This fraction (B) contained about half the galactose residues in the wood. Further extraction of the holocellulose successively with hot water, 14 % potassium hydroxide and 24 % potassium hydroxide containing 3 % boric acid² yielded fractions C, D and E, respectively. The results of these extractions are summarised in Table 1.

Fractions B and C were subfractionated by precipitation with barium hydroxide³, yielding precipitates B I and C I, and soluble fractions B II and C II. From analysis of hydrolysates of these fractions (Table 1) it was evident that B I was a fairly pure galactan containing 12.9 % uronic acids and contaminated with small amounts of glucomannan. This fraction, representing about 44 % of the galactose residues originally present in the wood, was used for the present investigation.

Arabogalactans have been isolated from several coniferous woods, including spruce wood⁴, and the possibility was considered that the present galactan might be derived from an arabogalactan, from which arabinose residues had been hydrolysed during delignification. It was found, however, that a larch

Table 1. Carbohydrate analyses of polysaccharide fractions from spruce compression wood.

	Yield g	Galactose %	Glucose %	Mannose %	Arabinose %	Xylose %
Spruce compression wood, extracted with acetone (Klason lignin: 38.8 %)	391.0	19.6	55.6	11.5	2.6	10.7
A: H ₂ O-extract	1.3	74.3	4.5	16.1	4.1	0.9
B: Polysaccharides from the delignification solution						
B I: Ba(OH) ₂ precipitate	21.4	97.3	trace	2.7	trace	trace
B II: Centrifugate	5.5	72.1	2.2	1.9	3.7	20.1
Holocellulose (Klason lignin: 1.1 %)	208.0	7.5	65.8	11.9	2.4	12.5
C: Polysaccharides from the H ₂ O extract of the holocellulose						
C I: Ba(OH) ₂ precipitate	5.4	45.2	11.6	41.3	1.0	0.9
C II: centrifugate	2.5	33.1	8.4	2.8	5.8	49.9
D: Polysaccharides from the 14 % KOH extract	40.7	19.0	5.2	12.4	6.2	57.2
E: Polysaccharides from the 24 % KOH + 3 % H ₃ BO ₃ extract						
E I: Ba(OH) ₂ precipitate	6.6	6.7	23.3	68.3	0.8	0.9
Residue		1.5	91.3	6.1	—	1.0

arabogalactan, after similar treatment, could be recovered unchanged. Furthermore the arabogalactans have highly branched structures, while the present polysaccharide gave a fairly strong film, characteristic for a polysaccharide with an essentially linear structure^{5a}. The infrared spectrum of the polysaccharide showed a strong band at 886 cm⁻¹, indicating a β -pyranosidic structure⁶.

Hydrolysis gave D-galactose and a number of acidic components, four of which were isolated in a fairly pure state by chromatography on thick filter paper. One of these acids was very probably galacturonic acid. A preliminary examination by paper chromatography and electrophoresis of hydrolysates of the other three acids indicated that in addition to galactose, one of them contained galacturonic acid and the other two glucuronic acid.

The possibility that part of the galacturonic acid originated from pectic acid was examined but attempted fractionation with cetyl-trimethylammonium hydroxide (CTA-OH), which is known to precipitate acidic polysaccharides⁷, was not successful. A spectrophotometric investigation of the polysaccharide revealed the presence of about 9 % lignin; by treatment with chlorine and extraction with ethanolic ethanolamine this value was reduced to 1 %. The delignified material on electrophoresis on glass paper gave a very elongated spot with M_G -value ranging from about 0.45 to 0.97. This indicated that the sample was highly polymolecular but no distinct components could be detected. Even after delignification the sample could not be separated by fractionation with CTA-OH into a neutral galactan and a pectic acid. This corroborates the results of the hydrolyses mentioned above, that showed that at least part

of the uronic acid is bound to the galactan. Whether these residues are present on the galactan chains as single residues or as chains of pectic acid type, as was recently suggested by Aspinall *et al.*⁸ for a galactan and a pectic acid from sisal hemp, could not be decided.

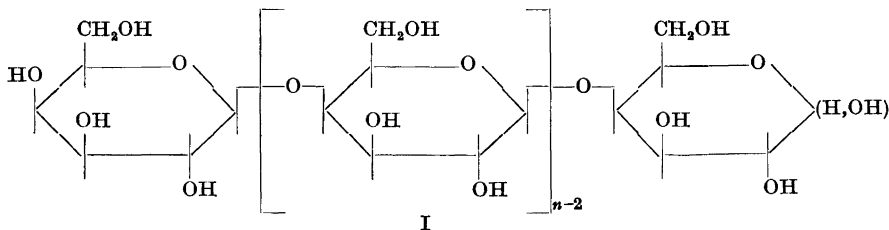
Since a nitrate ester of the galactan (N 11.8 %) was only partly soluble in acetone or butyl acetate, the osmotic pressure was determined on the sodium salt of the unsubstituted galactan in 0.1 N sodium chloride solution. The value obtained, \overline{DP}_n 52, is however rather uncertain since the sample also contained lignin.

Partial hydrolysis of the galactan and removal of the acidic components left a number of neutral sugars, which were fractionated by carbon column chromatography. The first four components were completely separated but only a partial separation was obtained of four more components which followed

Table 2. R_F (solvent b) and M_G (borate buffer, pH 10) values of the neutral components.

Fraction No.	Component	R_F	M_G
34—64	galactose	0.39	0.95
116—148	galactobiose	0.27	0.48
180—202	galactotriose	0.19	0.40
210—230	galactotetraose	0.14	0.35
232—246	galactopentaose	0.10	0.32
243—255	galactohexaose	0.07	0.31
253—270	galactoheptaose	0.05	0.30
262—281	galactooctaose	0.04	0.28

these. D-Galactose and a galactobiose were obtained crystalline. The galactobiose, $[\alpha]_D^{25} + 67^\circ$ showed the same paper chromatographic and electrophoretic data and the same X-ray diffraction pattern as the 4-O- β -D-galactopyranosyl-D-galactose isolated from white birch⁹, a sample of which was obtained by courtesy of Dr. T. E. Timell. Further its melting point 210—212° was undepressed on admixture with Timell's sample. The R_F and M_G values of the sugars are given in Table 2. The R_M -values, plotted against n , give a straight line (Fig. 1), indicating that the sugars belong to a homologous series of the general formula I.



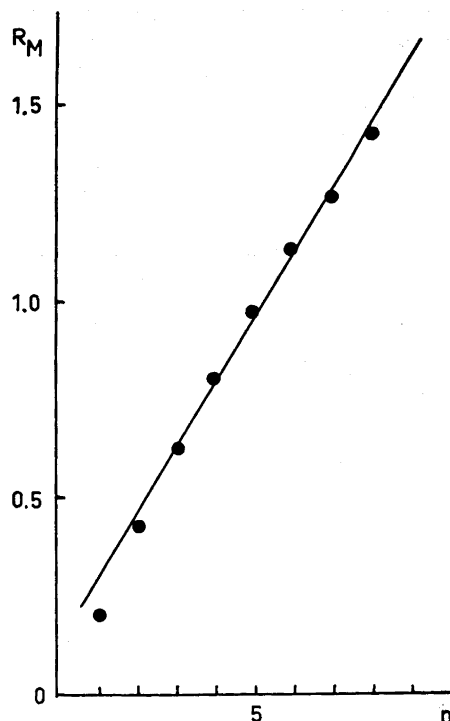


Fig. 1. Relation between R_M -values and chain length.

The electrophoretic data are also in agreement with this conclusion and each of the components on partial hydrolysis yielded all the lower oligosaccharides. Only traces of neutral oligosaccharides, other than those belonging to this series, were observed in the hydrolysate of the galactan, and the galactan must therefore be composed essentially of linear chains of β -1,4-linked D-galactose residues. Uronic acid residues are linked to some of these. The nature of these linkages and the question of whether some of the uronic acid residues are present in pectic acid chains must await further study.

The galactan from spruce compression wood is thus of similar type to that belonging to the pectic triad^{5b}. A similar galactan has recently been obtained by Andrews *et al.*¹⁰ from *Strychnos nux vomica* seeds and the isolation of 4- β -D-galactopyranosyl-D-galactose by Gillham and Timell¹¹ from a hydrolysate of white birch α -cellulose also indicates the presence of a polysaccharide of this type. The presence of pectic substances in the middle lamella of wood fibres has long been assumed, and it has recently been demonstrated^{12,13} that the middle lamella of unripe aspen fibres and of pine tracheids contain a high percentage of polymers, built up from galactose, arabinose and galacturonic acids, which are the constituent sugars of the pectic triad. Although a large part of the galactans from the compression wood was extracted during the delignification, a considerable part remained undissolved even after treat-

ment with 14 % potassium hydroxide. This does not necessarily imply that the wood contains galactans that are structurally very different. The same difference has also been observed in spruce glucomannans¹⁴, two extreme fractions of which showed some difference in DP but had essentially the same chemical structure.

EXPERIMENTAL

Melting points are corrected. Optical rotations were determined in water: c , 1.5. The concentration of solutions was done under reduced pressure, at a bath temperature of about 40°.

Paper chromatograms were run on Whatman No. 1 and 3 MM papers, using the solvent systems (v/v):

- a) Ethyl acetate-acetic acid-water, 3:1:3 (upper phase)
- b) Ethyl acetate-pyridine-water, 2:1:2 (upper phase).

The method of Saeman *et al.*¹⁵ was used for quantitative determinations. Paper electrophoreses were run on Schleicher and Schüll glass fibre papers and on Whatman No. 3 MM paper in 0.1 M borate buffer of pH 10 or 0.1 M acetate buffer of pH 4.

Isolation and fractionation of polysaccharides. Acetone-extracted wood meal (18–35 mesh) was extracted with hot water. Acidification and precipitation with ethanol, yielded fraction A. The wood meal was then delignified by the chlorite method at 60° and pH 4.7. The combined chlorite solutions were dialysed against running tap water for 11 days, acidified and precipitated with ethanol, yielding fraction B. The holocellulose was then extracted successively with hot water, 14 % potassium hydroxide and 24 % potassium hydroxide with 3 % boric acid. The latter two extractions were made under nitrogen, and the polysaccharides extracted were recovered by precipitation with ethanol (fractions C, D and E respectively).

Fractions B and C were dissolved in water and a saturated solution of barium hydroxide was added³. The material in the precipitates and in the supernatants was recovered, yielding fractions B I, C I and B II, C II respectively.

B I was delignified with chlorine and ethanolic ethanolamine, as described previously¹⁶. Treatment with cetyl-trimethylammonium hydroxide (CTA-OH) precipitated about 2/3 of the material. The precipitate and the material remaining in solution were isolated as described by Bouveng and Lindberg¹⁷. They had optical rotations of +80° and +55°, resp., the first fraction having a considerably higher content of uronic acid residues. Attempted fractionation of undelignified material with CTA-OH failed.

B I was readily soluble in water, $[\alpha]_D^{25} +57^\circ$. It contained about 9 % lignin¹⁸ and 12.9 % of uronic acid residues¹⁸, calculated as galacturonic acid. The \overline{DP}_n was determined osmotically as already described for an araboglucuronoxylan¹⁶.

Hydrolysis of B I. A solution of B I (3 g) in 4 % sulphuric acid (800 ml) was kept at 120° for 1 h and then neutralised with barium hydroxide. The neutral solution was passed successively through columns of Dowex 50 (H+) and Dowex 3 (free base) ion exchange resin. D-Galactose was recovered from the deionised solution. The acids were eluted from the anion exchange resin with 1 N ammonium hydroxide and the solution obtained was concentrated to a syrup. Electrophoresis and paper chromatography (solvent a) revealed the presence of galacturonic acid and of at least three acids with lower R_F values. The latter were separated by chromatography on thick filter paper. A chromatographic and electrophoretic examination of these acids after hydrolysis with 6 % sulphuric acid at 120° for 6 h indicated that one of them contained galactose and galacturonic acid while the other two were composed of galactose and, probably, glucuronic acid.

Partial hydrolysis of B I. A sample of B I (3 g) was heated with N sulphuric acid (200 ml) for 40 min on a steam bath, cooled and deionised. The solution was concentrated to 10 ml and then adsorbed on a carbon-Celite column (4 × 45 cm). Gradient elution with aqueous ethanol, 1 – 10 % (4 l) followed by 10–35 % (6 l) effected a separation of the components. The first four members, galactose to galactotetraose were completely separated. From the pentaose to the octaose they only partially separated, but relatively

pure top fractions were obtained of all components. The R_F - and M_G -values of the components are given in Table 2.

D-Galactose (346 mg), $[\alpha]_{25}^D +82^\circ$, m. p. 164–167°, undepressed on admixture of an authentic sample.

Galactobiose (278 mg), $[\alpha]_{25}^D +67^\circ$, m. p. 210–212°, alone or mixed with β -1,4-galactobiose, kindly supplied by Dr. T. E. Timell, Montreal. The X-ray diffraction pattern was identical with that reported by Gillham *et al.*⁹ The M_G -value in borate buffer, 0.48, was, as expected, lower than the values reported for the corresponding β -1,3- and β -1,6-linked disaccharides¹⁹ (0.69 and 0.84).

Galactotriose (213 mg), $[\alpha]_{25}^D +58^\circ$. Amorphous.

Galactotetraose (151 mg), $[\alpha]_{25}^D +53^\circ$. Amorphous.

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